Accepted Manuscript

International Journal of Sports Medicine

Innovations in the Assessment of Skeletal Muscle Health: A Glimpse into the Future

Jonathan P Beausejour, Kevan S Knowles, Abigail T Wilson, L. Colby Mangum, Ethan C Hill, William J Hanney, Adam J Wells, David H Fukuda, Jeffrey Stout, Matt S Stock.

Affiliations below.

DOI: 10.1055/a-2242-3226

Please cite this article as: Beausejour J P, Knowles K S, Wilson A T et al. Innovations in the Assessment of Skeletal Muscle Health: A Glimpse into the Future. International Journal of Sports Medicine 2024. doi: 10.1055/a-2242-3226

Conflict of Interest: The authors declare that they have no conflict of interest.

Abstract:

Skeletal muscle is the largest organ system in the human body and plays critical roles in athletic performance, mobility, and disease pathogenesis. Despite growing recognition of its importance by major health organizations, significant knowledge gaps remain regarding skeletal muscle health and its crosstalk with nearly every physiological system. Relevant public health challenges like pain, injury, obesity, and sarcopenia underscore the need to accurately assess skeletal muscle health and function. Feasible, non-invasive techniques that reliably evaluate metrics including muscle pain, dynamic structure, contractility, circulatory function, body composition, and emerging biomarkers are imperative to unraveling the complexities of skeletal muscle. Our concise review highlights innovative or overlooked approaches for comprehensively assessing skeletal muscle in vivo. We summarize recent advances in leveraging dynamic ultrasound imaging, muscle echogenicity, tensiomyography, blood flow restriction protocols, molecular techniques, body composition, and pain assessments to gain novel insight into muscle physiology from cellular to whole-body perspectives. Continued development of precise, non-invasive tools to investigate skeletal muscle are critical in informing impactful discoveries in exercise and rehabilitation science.

Corresponding Author:

Dr. Matt S Stock, University of Central Florida, Institute of Exercise Physiology and Rehabilitation Science, Orlando, United States, matt. stock@ucf.edu

Affiliations:

Jonathan P Beausejour, University of Central Florida, Institute of Exercise Physiology and Rehabilitation Science, Orlando, United States Kevan S Knowles, University of Central Florida, Institute of Exercise Physiology and Rehabilitation Science, Orlando, United States Abigail T Wilson, University of Central Florida, Institute of Exercise Physiology and Rehabilitation Science, Orlando, United States [...]

Matt S Stock, University of Central Florida, Institute of Exercise Physiology and Rehabilitation Science, Orlando, United States

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



Downloaded by: Harriet F. Ginsburg Health Sciences Library. Copyrighted material.

- Innovations in the Assessment of Skeletal Muscle Health: A
 Glimpse into the Future
 3
- 4 Running Title: Innovations in Skeletal Muscle Assessment
- 5 Word Count: 5,286
- 6 Figures: 7
- 8 Abstract

Skeletal muscle is the largest organ system in the human body and plays critical roles in athletic performance, mobility, and disease pathogenesis. Despite growing recognition of its importance by major health organizations, significant knowledge gaps remain regarding skeletal muscle health and its crosstalk with nearly every physiological system. Relevant public health challenges like pain, injury, obesity, and sarcopenia underscore the need to accurately assess skeletal muscle health and function. Feasible, non-invasive techniques that reliably evaluate metrics including muscle pain, dynamic structure, contractility, circulatory function, body composition, and emerging biomarkers are imperative to unraveling the complexities of skeletal muscle. Our concise review highlights innovative or overlooked approaches for comprehensively assessing skeletal muscle in vivo. We summarize recent advances in leveraging dynamic ultrasound imaging, muscle echogenicity, tensiomyography, blood flow restriction protocols, molecular techniques, body composition, and pain assessments to gain novel insight into muscle physiology from cellular to whole-body perspectives. Continued development of precise, non-invasive tools to investigate skeletal muscle are critical in informing impactful discoveries in exercise and rehabilitation science.

Keywords: pain; injury; rehabilitation; athlete performance; blood flow restriction; muscle quality

45 Introduction

46

47 Skeletal muscle is one of the most metabolically active and adaptable tissues in the human body,

48 comprising up to 40% of total body mass and containing 50-75% of all body proteins [1]. The

49 dynamic and plastic nature of skeletal muscle enables it to support a wide range of vital functions

for human health and performance, including initiating movement, maintaining posture and body temperature, stabilizing joints, and storing nutrients [2]. Over recent decades, researchers have

52 developed and refined a range of techniques to evaluate skeletal muscle structure and function,

53 non-invasively. These include imaging modalities such as magnetic resonance imaging (MRI)

and CT, dual energy X-ray absorptiometry, anthropometric measurements such as skinfolds and girths, electromyography (EMG), and isokinetic dynamometry. Application of these techniques

has provided key insights into the adaptability of human skeletal muscle within the context of

aging, disease, injury, exercise, and nutrition [1,3,4]. While current methods have advanced our

58 understanding of skeletal muscle physiology, continued innovation and optimization are

necessary to develop more feasible assessment tools capable of exploring intricate muscle
morphology responses to different physiological and pathophysiological stimuli [5]. Emerging

morphology responses to different physiological and pathophysiological stimuli [5]. Emerging
areas requiring further research include the influence of individual variation in muscle structure
and function, sensitivity of assessment techniques, the interplay between muscle and other

63 tissues like fat and bone, and the ideal modes and dosages of exercise, nutrition, and

64 rehabilitation interventions [6,7].

65 66 To further promote engagement in these research avenues, scientists must continue honing 67 current approaches while implementing more viable, novel assessment tools aimed to adequately assess skeletal muscle properties. The purpose of our review is to briefly highlight emerging, 68 innovative, and relatively feasible approaches that show promise in assessing skeletal muscle 69 70 health. Covered topics in this review include ultrasound-derived dynamic imaging, tensiomyograpy, innovative approaches for blood flow restriction administration, utilization of 71 72 neoepitope-biomarkers for skeletal muscle structure and function, ultrasound-derived echo 73 intensity measures for muscle quality, and other novel assessments of body composition and 74 skeletal muscle pain.

76 Methods

A comprehensive review of the science literature was conducted to assess the latest approaches
in evaluating skeletal muscle health. The National Institutes for Health National Library of
Medicine (PubMed.gov) and Google Scholar search engines were utilized, specifically, to
identify recent publications related to dynamic imaging of skeletal muscle, peripheral
neuromuscular assessments, muscle echogenicity, blood flow restriction, biomarkers of skeletal
muscle function, skeletal muscle quality, body composition, and pain assessment.

84

75

85 **Dynamic Imaging**

86

87 Muscle function and characteristics can be measured in several ways and many laboratory- and

- 88 hospital-based studies utilize EMG, and imaging instrumentation, such as magnetic resonance
- 89 imaging. Financial and time constraints can limit access and the ability to employ many of these
- 90 techniques when clinicians and practitioners are seeking a deeper understanding of muscle

- function. A viable alternative to some of the more expensive and less accessible options is
 ultrasound imaging (Fig 1) [8]. Specifically for musculoskeletal ultrasound imaging, there are
 established methods detailing how to measure muscle thickness, cross-sectional area,
 echogenicity, etc. [9]. Most of these methods rely on the patient or participant to be in a static,
 rested state, which controls the environment with hopes to avoid any artifact or false
- representation of the muscle's morphology. However, many of the patient populations included
- in ultrasound imaging studies and clinical scenarios are likely to experience some sort of injury
- and/or exhibit symptomology that revolves around pain during movement.

99 .00 Ultrase

Ultrasound imaging when used in a dynamic, innovative manner, capturing images during 100 101 contraction and even exercise, provides an understanding of muscle function, not just static 102 characteristics [8]. When a patient or participant is experiencing pain and dysfunction during movement, activities of daily living, or exercise, the consequences are well documented, 103 especially in those with chronic low back pain. Due to the known contribution of abdominal, hip, 104 and pelvic muscles to low back pain-related dysfunction, ultrasound imaging is a useful tool to 105 view the complex layering of those muscles [8]. Part of the injury assessment process could 106 107 include viewing muscles when individuals move, and they experience pain or avoid movement due to the fear of pain. Capturing muscle thickness, cross-sectional area, and quality can be done 108 reliably in many positions [9,10], commonly in the lateral abdominal wall and posterolateral hip. 109 110 The application of dynamic ultrasound imaging has been described recently for sport-specific and body part-focused rehabilitation exercises [11,12]. Visualizing musculature of the lateral 111 abdominal wall is possible by fixing the ultrasound probe to the anterolateral abdomen with a 112 113 belt ensuring the probe stays in the same position even while the individual moves. Documented methods describing this technique include capturing images and videos while walking, 114 balancing, squatting, planking, swinging a golf club, etc.[11–14].]. Populations included in these 115 116 studies span from healthy, asymptomatic individuals to those experiencing low back pain and chronic ankle instability. 117

118 The utility of dynamic ultrasound imaging is shown by recent studies that have reported muscle 119 120 thickness changes from a static, rested position to an active, contracted position. Activation ratios 121 may also be calculated by dividing the contracted thickness by the rested thickness and were first established in hook-lying tabletop positions. Functional activation ratios divide thickness during 122 123 exercise (e.g., peak knee flexion during a single leg squat) by a static, starting position (e.g., 124 standing). A preferential activation ratio [15] involves comparing the thickness of one muscle to others within the same image. For example, when imaging the lateral abdominal wall, the change 125 in thickness of the transverse abdominis could be divided by the change in thickness of the entire 126 127 lateral abdominal wall (Figure 2). This preferential ratio provides insight into how much the 128 transverse abdominis changes its thickness relative to the other muscles during contraction. A 129 greater preferential activation ratio indicates the transverse abdominis is the predominant muscle changing thickness out of the entire lateral abdominal wall. Dynamic ultrasound imaging also 130 allows for an innovative, clinical approach to visual biofeedback. As the patient visualizes their 131 132 muscles during a prescribed exercise, activity of daily living, or pain-provoking position, there is 133 an opportunity to show the patient how they can contract the muscle either at a different time or how to increase thickness in general. Methods have been established for sport-specific 134 135 ultrasound biofeedback, specifically viewing the obliques during a golf swing [12]. Brightness, 136 B-mode, and Motion, M-mode can be used for dynamic imaging and biofeedback. Based on the

- goal of dynamic imaging, B-mode may apply when viewing a reference, static image, or to view
 a frame-by-frame breakdown of a muscle moving through a task. M-mode may be appropriate
 when synchronizing dynamic ultrasound with other muscle measurement tools capturing in the
 time domain, such as timing of activation with electromyography [11].
- 141

142 Contractile Properties

143

144 Tensiomyography (TMG) is a non-invasive and objective assessment tool used to evaluate skeletal muscle contractile properties and peripheral neuromuscular function. It provides 145 valuable information about muscle contraction characteristics, muscle fiber composition, and 146 147 muscle fatigue. TMG has gained popularity in sports science, rehabilitation, and research settings 148 due to its ability to provide real-time and reliable data on muscle function [16-18]. TMG involves the application of an external electrical stimulus to the muscle belly, causing a muscle 149 twitch response that is measured through a displacement sensor tip (Fig 2). It primarily assesses 150 151 muscle contractile properties, including muscle displacement, contraction time, and muscle relaxation time. These parameters reflect muscle stiffness, contractile speed, and muscle fiber 152 153 recruitment patterns [19,20]. One of the key advantages of TMG is its ability to provide objective and quantitative data. Traditional assessment methods, such as manual muscle testing 154 or electromyography, often rely on more subjective, post-acquisition interpretation or qualitative 155 156 measures. TMG, on the other hand, offers standardized numerical values and objective 157 measurements, reducing the potential for human error and enhancing the reliability of the 158 assessment [20,21]. By tracking changes in muscle contractile properties, one can evaluate the 159 effectiveness of training programs and make adjustments accordingly. TMG can also help 160 identify muscle imbalances and guide targeted interventions to restore balance and optimize 161 performance.

163 TMG involves elicited, involuntary isometric contractions that are generated using a single 1 mswide biphasic wave. By utilizing proprietary computer software, a twitch curve is generated 164 based on data from the sensor, which allows for the determination of six primary parameters. 165 166 The y-axis of this curve represents muscle displacement in millimeters, while the x-axis represents time in milliseconds. The key TMG parameters comprise displacement (Dm), 167 contraction time (Tc), delay time (Td), contraction velocity (Vc) (Vc=[90%Dm-10%Dm/Tc]), 168 169 sustain time (Ts), and half-relaxation time (Tr). (6) Displacement (Dm) pertains to the highest radial displacement achieved by the muscle and is linked to muscle stiffness. Contraction time 170 (Tc) represents the duration between 10% and 90% of Dm on the positive slope of the twitch 171 172 curve. Delay time (Td) is a temporal parameter that measures the duration from the initiation of 173 the electrical stimulus to when the muscle belly reaches 10% of Dm or peak displacement. Half-174 relaxation time (Tr) refers to the time taken for the muscle displacement to decrease from 90% of 175 its maximum to 50% of Dm on the negative slope of the curve. Sustain time (Ts) is defined as 176 the time between 50% Dm on both the negative and positive slopes of the curve. Contraction velocity (Vc) is a calculated metric that aims to quantify the rate of muscular contraction. Since 177 178 Vc is a derived measure, various methods have been employed by authors to compute it. The 179 most commonly used calculation involves dividing the change in Dm between 10% and 90% by 180 Tc. This approach enhances the usefulness of the Tc parameter and provides a more reliable 181 measure of contraction speed by mitigating the influence of Dm. This is important as peak radial

displacement values have been shown to impact contraction time values due to the inherentshape of the twitch curve [20,21].

184

202

185 TMG can also help identify specific muscle deficits or imbalances that may contribute to functional limitations or recurring injuries. With this information, tailored rehabilitation 186 programs can be developed to address these deficits and promote optimal recovery [18,21]. 187 Moreover, TMG can be a valuable research tool for investigating muscle adaptations to training, 188 189 comparing different training protocols, or studying the effects of injury or disease on muscle 190 function. Researchers can use TMG to examine changes in muscle contractile properties following specific interventions or to explore how different training modalities affect muscle 191 192 performance. 193

While TMG has shown great promise as an assessment tool, it is important to note its limitations.
TMG primarily focuses on muscle contractile properties and does not provide direct information
about neural activation or muscle force production. It is also important to consider that TMG
measurements may be influenced by factors such as skin impedance, adipose tissue thickness,
and anatomical variations. These factors need to be considered during data interpretation to
ensure accurate and meaningful results.

201 Innovation Applications to Determine Blood Flow Restriction Occlusion Pressures

203 Traditional exercise-based approaches are not well tolerated by several clinical populations, an 204 issue researchers/practitioners continually attempt to circumvent [22–24]. An exciting strategy to 205 address this issue includes combining exercise with limb occlusion (blood flow restriction [BFR]) due to its ability to induce similar or even superior benefits compared to traditional non-206 207 occluded exercise [25-27]. During BFR exercise, low exercise loads (e.g., lighter weights/resistance) are used while completing a standardized 5-min exercise scheme. Blood flow 208 restriction exercise uses a small inflatable cuff applied to the upper most portion of a limb to 209 restrict venous blood from exiting the exercising limb which facilitates robust physiological 210 211 responses that may underly the subsequent increases in muscle strength, mass, and endurance 212 (i.e., fatigue resistance) observed following chronic BFR exercise. These adaptations have been demonstrated in asymptomatic [22–31] and some symptomatic populations [32–34]. Importantly, 213 214 BFR resistance exercise has also been shown to be safe and to elicit positive effects in post-215 surgical [35,36] older adults [32,37,38] and hospitalized patients [39]. Specifically, the safety and effectiveness of BFR exercise has been routinely demonstrated with the implementation of 216 217 the standard 75 repetition $(1 \times 30, 3 \times 15)$ scheme [28,40]. Additionally, the exercise is 218 performed at 30% of 1RM (i.e., a low weight/resistance) and with the small, inflated cuff set to a 219 pressure corresponding to 40-80% of arterial occlusion pressure (i.e., approximately 40-80% of 220 systolic blood pressure) [41,42].

221

Despite its high versatility and implementation in both research, clinical, and athletic settings, the application of BFR is limited. Specifically, the gold standard for the determination of total arterial occlusion pressure (TAOP) and application of BFR requires the use of pulsed wave Doppler (Fig 3). Typically, this necessitates trained personnel to operate and interpret ultrasound-based readings (i.e., arterial and venous blood flow) with the concomitant modulation of pneumatic pressures. Theoretically, minute variations in the determination of TAOP changes

the BFR pressure applied which has been demonstrated to alter the physiological responses 228 229 [41,43] and discomfort (e.g., higher pressures result in greater discomfort). Therefore, it is imperative that TAOP is determined accurately and precisely. As a result, the current 230 231 applications of BFR exercise are somewhat limited to research and clinical settings whereby these procedures can be performed accurately and reliably, although data are limited in this 232 regard. For example, among a small sample of males (n=13), pulsed wave Doppler ultrasound 233 exhibited moderate/high reliability (intraclass correlation coefficient [ICC] = 0.796), while the 234 235 coefficient of variation (COV) was 5.6% (Bezerra et al., 2017). There were, however, no relative measures of reliability reported (e.g., standard error of the measurement [SEM] or minimal 236 237 difference [MD]) which limits the application of these findings [44]. 238

239 An exciting strategy to circumvent the determination of TAOP, is the algorithm-based determination of TAOP. Specifically, several commercially available devices (e.g, SujiBFR, 240 Defli, Smartcuffs) independently estimate TAOP using proprietary engineering that likely 241 242 leverages known variables (e.g., limb width, mean arterial pressure) which affect TAOP. For example, in a small sample (n=10), one study [45] examined test-retest reliability of an 243 244 algorithm-based determination (Delfi PTS, Delfi Medical, Vancouver, BC, Canada) of TAOP and reported high reliability (ICC > 0.953; COV = 2.97%), but the validity of this device was not 245 examined. Algorithm-based devices such as the one previously examined and others (e.g., Suji, 246 247 SmartCuffs) may exhibit greater utility than current research-based practices for the 248 determination of TAOP which requires ultrasound and trained personnel, but their algorithms 249 should be critically examined to prevent potential adverse effects. Specifically, researchers 250 [42,46,47] have done substantial and meaningful work identifying predictors, equations, and/or 251 algorithms which can estimate TAOP, but these methods are not without error. Thus, there is an inherent need to determine if commercially available BFR devices implementing algorithms to 252 determine TAOP are clinically sound (i.e., reliability and validity). Furthermore, there is 253 insufficient data examining the reliability of TAOP using pulsed wave Doppler (the criterion 254 method). Regardless, commercially available devices (e.g., SujiBFR, Delfi, Smartcuffs) may 255 256 provide viable alternatives to the criterion determination of TAOP ultimately facilitating the 257 larger implementation of BFR. 258

Novel Serological Neoepitopes as Biomarkers of Skeletal Muscle Structure and Function 260

Biomarkers are defined as measurable indicators of biological processes or responses to an 261 exposure or intervention [48]. They represent an indispensable tool in human biology that 262 263 allows researchers to map the complex physiological pathways that underpin healthy and altered physiological function [49]. The use of serological biomarkers has the benefit of being relatively 264 265 non-invasive and easy to perform [50,51], which has led to the widespread development of 266 assays for various biological markers. In this regard, neoepitopes have emerged as a class of 267 serological peptide biomarkers with diagnostic and prognostic potential that may also have utility as minimally invasive indicators of health status, physiological response and/or disease 268 269 susceptibility (Fig 4). 270

271 In skeletal muscle, neoepitopes may be exposed following post-translational modifications

272 (PTMs) to specific muscle proteins [52]. Proteolytic cleavage is a particularly interesting PTM

273 as it creates peptide fragments and the potential for novel neoepitopes on the carboxy- or 274 aminoterminal ends of cleaved peptides [53,54]. Peptide fragments being smaller than their intact 275 parent proteins can enter the circulation more readily [55] where antibodies can be raised against 276 specific neoepitopes exposed on these fragments. Since serological neoepitope biomarkers 277 consist of a unique combination of parent proteins and PTMs [56], they are thought to be 278 reflective of tissue specific physiological or pathological remodeling processes rather than 279 overall muscle size or quality and may therefore serve as ideal biomarkers for early the detection 280 of various myopathies [57,58]. It is also thought that serological neoepitope biomarkers have the 281 potential to provide insight into net changes in protein metabolism, which is currently limited to 282 operationally complex and invasive stable isotope techniques [58].

283 284 Much of the research examining neoepitopes in skeletal muscle has focused on temporal changes to extracellular matrix (ECM) collagens following various interventions. Nedergaard et al., 2013 285 286 examined changes in collagen type VI fragment degraded by matrix metalloproteinases 2 and 9 287 (C6M) and type VI collagen N-terminal globular domain epitope (IC6) among young and old 288 men at baseline after 2-weeks of unilateral immobilization and 4-weeks of remobilization with 3 289 x weekly resistance training, respectively [59]. They found significant correlations between IC6 and muscle mass at baseline, and between C6M and the change in muscle mass from 2-weeks to 290 291 4-weeks of remobilization in young but not old men [59]. The same group also reported significant associations between lean body mass, IC6, collagen type III synthesis (Pro-C3) and 292 the IC6/C6M ratio among matched controls in the 25B cohort of the Danish Head and Neck 293 294 cancer group (DAHANCA) trial [60]. In another study, Sun et al., 2015 examined the temporal 295 profile of neoepitope peptides Pro-C3, C-terminus α3(VI) chain (Pro-C6) and C6M following 8-296 weeks immobilization and remobilization. They reported significant associations between Pro-C3, C6M and lean body mass at baseline, a significant upregulation of both Pro-C3 and Pro-C6 297 298 following immobilization and remobilization indicative of muscle remodeling, and an inverse 299 relationship between Pro-C6 and changes in muscle mass [61]. Consistent with this, work by Nielsen et al. 2013 also indicates that higher levels of Pro-C3 predict greater muscle mass in 300 healthy individuals [62]. More recently, Reule et al., 2016 found that the ratio of type II collagen 301 302 collagenase cleavage neoepitope (C2C) to C propeptide of type II procollagen (CP2) was 303 responsive to leucine-rich amino acid supplementation administered in conjunction with 12-304 weeks of combined aerobic strength and balance training [63]. They report a significantly greater decrease in the acute phase (0-3 hours) C2C/CP2 post-training response to a downhill walking 305 306 stress test when compared to the placebo group, indicating a lower disturbance in joint homeostasis that coincided with a significant attenuation of acute phase quadricep MVC strength 307 308 loss [63]. Several other serological biomarkers, including C-terminal agrin fragment (CAF) and matrix metalloproteinase-2 degraded titin fragment (titin-MMP2) also appear to be strong 309 discriminators of normal versus aberrant skeletal muscle outcomes including muscle 310 311 wasting/atrophy and protein turnover [61,64].

312

The large dynamic range and complexity of both the proteome and various PTMs [65] represents a significant challenge for the identification of new muscle specific neoepitope biomarkers and

315 targeting reagents. Nevertheless, while this area of research is currently in its infancy, the limited

- 316 current literature indicates that serological neoepitope biomarkers of cleaved circulating peptide
- 317 fragments are promising candidates for assessing skeletal muscle structure and function.

318319 Skeletal Muscle Quality

320

329

321 Studies have increasingly shown that there is a disassociation between skeletal muscle strength and mass [66,67]. For example, the National Institutes on Aging's longitudinal Healthy, Aging, 322 323 and Body Composition study showed that adults \geq 70 years of age lost \sim 3× more muscle strength than mass on an annual basis [66]. Even more compelling, older adults that gained muscle mass 324 325 still lost muscle strength [68]. In addition, immobilization and bed rest studies have shown that muscle strength and mass show divergent timelines, with strength rapidly diminishing before 326 327 detectable declines in muscle mass [69–71]. These concepts have given rise to the concept of 328 muscle quality and its methods of assessment [72].

330 The measurement of echo intensity has emerged as a potential tool for studying skeletal muscle 331 quality and estimating intramuscular fat content, providing insights that may be unique from measures of muscle size [73]. Echo intensity is a quantitative measure of brightness in ultrasound 332 images, reflecting the echogenicity of tissues (Fig 5). In skeletal muscle, echo intensity is 333 334 influenced by physiological factors such as muscle fiber arrangement [74], connective tissue content [75], and the presence of intramuscular fat [76]. Higher echo intensity values are 335 associated with muscle pathologies like atrophy, fibrosis, and fatty infiltration [77], while lower 336 values are observed in young, healthy muscles. MRI studies suggest that echo intensity's ability 337 338 to estimate intramuscular fat content appears promising[77,78]. Echo intensity is associated with 339 several functional outcomes [79,80] and it appears to be sensitive in detecting differences 340 between age groups [81]. Additional research is needed to understand why large differences in echo intensity are often seen when comparing groups with distinct characteristics like age or 341 training status, whereas smaller changes are detected in response to exercise or rehabilitation 342 interventions [82]. One potential explanation is that intrinsic physiological factors like muscle 343 fiber type distribution, connective tissue content, and intramuscular fat infiltration change slowly 344 over time. Group differences may represent the cumulative result of prolonged exposure to 345 factors like aging. In contrast, short-term interventions elicit more modest echo intensity 346 347 changes, as muscle structural characteristics do not radically transform within days or weeks. Longer training studies are needed to determine if more marked echo intensity changes can be 348 induced over time with sustained exposure to stimuli like exercise. It is also important to 349 recognize that echo intensity is affected by methodological factors, such as probe tilt [83] and 350 participant positioning [84]. Recent evidence suggests that researchers new to echo intensity 351 352 analyses provide reliable measurements [85] and small adjustments in image depth to 353 accommodate muscles of different sizes are acceptable [86].

354

Most published echo intensity studies have relied on ImageJ software (National Institutes of
Health, Bethesda, MA, USA), in which investigators manually analyze a muscle's region of
interest. While these approaches are well established, they can be time consuming and
subjective, making it difficult to conduct large-scale analyses quickly. However, emerging
technologies are likely to streamline the measurement process, reduce subjectivity, and enhance
the accuracy of echo intensity analysis. Automated or semi-automated region of interest selection
algorithms have recently been introduced to target specific muscle regions [87,88]. In the future,

- 362 computerized analysis of ultrasound images will enable the precise quantification of echo
- intensity values, ensuring consistent and reproducible evaluations. In addition, small probes and
- 364 wireless technology that integrate with laptops, tablets, and smartphones phones will likely make
- the analysis of echo intensity much more accessible and rapid.
- 366

374

375

367 Overall, the measurement of echo intensity has emerged as an innovative approach for studying368 skeletal muscle quality and estimating intramuscular fat content. Importantly, these

369 measurements can be done with ultrasound devices that are portable and less expensive than

370 MRI, and minimal training is required. These measurements are particularly useful when

371 complementing measures of muscle size (e.g., cross-sectional area or volume) and physical

372 function. For more detailed information, the reader is encouraged to review two recent echo

intensity reviews by Stock and Thompson [73] and Wong et al [82].

Practical Body Composition & Novel Use of Bioelectrical Impedance Analysis (BIA)

Laboratory-based methods of body composition estimation potentially offer an error reduction of
approximately 50% in body fat estimation compared to field-based methods. However, these
improvements are partially attributable to the ability to control numerous physiological
assumptions [89]. The inherent error of body composition measurement is further complicated
using either population-specific or generalized prediction equations that likely compound the
error in some individuals or groups. For athletes, the need for dietary restriction, adequate
hydration, and standardized physical activity further exacerbates the issue.

384 385 Whole body estimation of body fat percentage is limited with its greatest utility in large population/epidemiological evaluations or public health settings. Site- or region-specific values 386 387 to evaluate tissue distribution may be more useful for practitioners and athletes. Dual-energy Xray absorptiometry enables potential evaluation of total body and site-specific fat mass, lean soft 388 tissue mass, and bone mineral density [89]. However, these devices are expensive, vary across 389 390 manufacturers, require low-dose radiation exposure, all of which may be prohibitive in many 391 practical settings. 392

A viable alternative is the direct evaluation of skinfold thickness either at a specific site or as a
sum of several sites throughout the body, instead of using prediction equations to determine body
density. While standardized protocols and trained evaluators are required, this method seems
least affected by the inherent variation in the typical restrictions recommended for body
composition assessments [90]. Although less commonly reported in the literature, recent
attempts have been made to provide normative data for the summation of skinfold measurements
[90,91].

400

As skinfold thickness does not directly assess skeletal muscle, it is recommended to evaluate this
information alongside anthropometric data such as regional circumferences (limb, hip/waist, etc.)
and/or body mass [92,93]. Fig 6 illustrates how changes in skinfold thickness can be translated to
body fat, while circumferences or body mass can serve as a proxy for muscle mass and the
interaction between these values.

407 This combined information can also be used to calculate corrected girth values (which account 408 for an adjustment of skinfold thickness to determine musculoskeletal cross-sectional area) or lean mass index (a log-based adjustment for body mass and summed skinfolds) [94]. Notably, 409 410 DeFreitas et al. [95] demonstrated that, despite underestimating muscle cross-sectional area compared to peripheral quantitative computed tomography (pQCT), both a corrected girth 411 equation [96] and a regression equation [97] adequately tracked changes in this value during an 412 eight-week resistance training program. A recent review by Duarte et al. [98] discusses 413 414 numerous validated anthropometric equations for limb-specific muscle mass estimation. 415 BIA offers a unique approach for body composition assessment by estimating the fat-free body, 416 417 generally assumed to be ~73% hydrated, rather than fat mass [89]. However, the basic 418 assumptions of body shape (i.e., the segments of the body are perfect cylinders), as well as the use of prediction equations (either through published work or developed by the device 419 manufacturer), introduce similar problems to other measurement methods. Therefore, it is 420 421

421 becoming increasingly common to directly evaluate the raw bioelectric parameters [resistance422 (R), reactance (Xc), and phase angle (PhA)] recorded by these devices (typically at 50 kHz).

423 424 From a practical perspective, R may represent cellular hydration, Xc may represent cell membrane integrity, and PhA is calculated as the arctangent ratio of Xc to R. The latter variable 425 426 may be considered representative of intra- and extra-cellular fluid (or ICW/ECW ratio) and/or 427 cell body mass, which has been suggested as an indicator of cellular health. These values can be 428 considered separately or plotted together through bioelectrical impedance vector analysis 429 (BIVA). The resultant vectors have been shown to differentiate between competitive levels and 430 types of athletes [99]. Interestingly, Kim et al. [100] demonstrated the discriminative potential of BIVA by distinguishing between female fashion models, dancers, and gymnasts in a manner 431 432 similar to somtatotyping with increasing mesomorphy (i.e., muscularity) and decreasing ectomorphy (i.e., linearity) across these groups. Furthermore, a recent systematic review reported 433 434 that PhA increases with age and is higher in athletes than controls as well as in males than 435 females [101].

The evaluation of raw bioelectrical data can also be applied regionally, a process known as
electrical impedance myography or localized BIA. Initially developed to examine diseased tissue
and subsequently, sarcopenic individuals, its application in athletic populations has been
established, focusing on adaptations to exercise and return-to-play situations following injury
[102].

In adopting an "innovation through simplification" stance towards body composition assessment,
this section of the paper underscores the importance of understanding and contextually applying
the available methodologies. For a comprehensive overview of available methods, and a practical
decision-making tree readers are encouraged to consult Kasper et al. [90]. Further, a detailed
discussion on related topics is provided by Lukaski & Raymond-Pope [99].

448

436

449 Skeletal Muscle Pain

450

451 Pain is a sensory and emotional experience impacted by the interaction of biological and

452 psychosocial factors [103]. One biologic factor that impacts the perception of pain is skeletal

453 muscle health. In a state of inflammation or musculoskeletal pathology, myofascial trigger points 454 may develop within the tissue that contribute to a myofascial pain syndrome [104]. Myofascial pain is prevalent with 30% of individuals seeking care for pain at a primary care office meeting 455 the criteria for myofascial pain. While the definition of a trigger point can vary, there is general 456 457 agreement among clinicians and scientists that myofascial pain is a separate diagnosis from fibromyalgia [105] and that trigger points contribute to myofascial pain syndrome [106]. 458 459 460 Myofascial trigger points are localized, taut bands of skeletal muscle tissue (Fig 7). A recent Delphi survey established that two of the following three criteria should be present to diagnose a 461 myofascial trigger point: taut band, hypersensitive spot, and referred pain [107]. During direct 462 463 compression, a 'jump response' may be elicited with or without referred pain [106]. Myofascial

trigger points are often both painful to palpation and can be a generator of pain. While the mechanisms underlying trigger points are multifactorial, repetitive eccentric contractions or overuse may result in an abnormal increase in acetylcholine at the neuromuscular junction of the muscle. Abnormal acetylcholine release may generate a sustained muscle contraction, causing localized ischemia and a palpable taut band [108]. Although myofascial pain was traditionally thought to only involve peripheral changes, muscle sensitization is also impacted by central nervous system sensitization [109].

471 472 A diagnosis of myofascial pain syndrome relies on the palpation of myofascial trigger points. 473 However, a limitation in the clinical assessment is varied inter-rater reliability in myofascial 474 trigger point identification [110]. New approaches have been proposed to improve the 475 identification of trigger points, including: pressure algometry and imaging. Pressure Pain Threshold (PPT) is a cost-effective and clinically feasible technique that may be employed to 476 477 assess trigger points. During PPT, a device with a small rubber tip (algometer) is applied over the 478 muscle with an ascending intensity until the individual reports that the sensation changed from 479 "comfortable pressure to slightly unpleasant pain" (pain threshold). The benefit of this 480 assessment is the stimulus is quantifiable and, therefore, the threshold for pain perception in 481 response to pressure is measured. PPT is significantly lower over trigger points and increases in 482 areas without trigger points [111]. Excellent intra and inter-rater reliability is demonstrated for 483 PPT application over trigger points [112].

485 Recent advances have also allowed for imaging of trigger points [113]. Imaging methods, including ultrasound, magnetic resonance imaging, and infrared thermography have been 486 developed as objective measures to potentially address the limitations with reliability. Imaging 487 allows for objective characterization of the tissue consistent with the definition of a trigger point. 488 489 B-mode ultrasound imaging indicates trigger points present as spherical, hypoechoic regions 490 [56]. Ultrasound elastography indicates decreased vibration amplitudes within the region, 491 indicative of localized stiffness of the muscle [114] at the site of the trigger point. Blood flow to 492 myofascial trigger points is also distinct from healthy tissue [114]. Magnetic resonance imaging has been used to examine trigger points; however, the evidence remains unclear on the benefit of 493 494 this imaging modality [115] for this purpose. Trigger points are important contributors to 495 myofascial pain and relevant to clinical treatment of patients with myofascial pain. Innovations 496 in standardizing the definition of trigger points, along with advances in the imaging of muscle, 497 may help to make this phenomenon more objective.

498

499 Summary

500

511

513

Accurate and precise assessment of skeletal muscle health is imperative for diagnosis of disease 501 502 and optimization of exercise and rehabilitation interventions. Our review has highlighted several viable, novel techniques with potential to advance these aims. Although not comprehensive, we 503 have focused on select emerging approaches based on their promise for impactful discoveries 504 505 and feasible implementation. Rapid technological innovations and subsequent adoption seem 506 poised to accelerate and expand prior methods. Despite progress, outstanding questions remain 507 regarding individual variation in exercise responsiveness, organ crosstalk, biomarker 508 development, and monitoring and enhancing athletic performance. It is our hope that continued 509 technical advances in assessing skeletal muscle health will provide insights into these critical 510 topics in exercise and rehabilitation science.

512 **References**

- 514 [1] Frontera WR, Ochala J. Skeletal muscle: a brief review of structure and function. Calcif
 515 Tissue Int 2015; 96: 183–195. Doi:10.1007/s00223-014-9915-y
- 516 [2] mcculler C, Jessu R, Callahan AL. Physiology, Skeletal Muscle. In: statpearls. Treasure
 517 Island (FL): statpearls Publishing; 2023
- 518 [3] Goodpaster BH, Carlson CL, Visser M, et al. Attenuation of skeletal muscle and strength in
 519 the elderly: The Health ABC Study. J Appl Physiol (1985) 2001; 90: 2157–2165.
 520 Doi:10.1152/jappl.2001.90.6.2157
- 521 [4] English KL, Paddon-Jones D. Protecting muscle mass and function in older adults during
 522 bed rest. Curr Opin Clin Nutr Metab Care 2010; 13: 34–39.
 523 Doi:10.1097/MCO.0b013e328333aa66
- [5] Raue U, Slivka D, Minchev K, et al. Improvements in whole muscle and myocellular
 function are limited with high-intensity resistance training in octogenarian women. J Appl
 Physiol (1985) 2009; 106: 1611–1617. Doi:10.1152/japplphysiol.91587.2008
- 527 [6] Timmons JA. Variability in training-induced skeletal muscle adaptation. J Appl Physiol
 528 (1985) 2011; 110: 846–853. Doi:10.1152/japplphysiol.00934.2010
- 529 [7] Churchward-Venne TA, Burd NA, Phillips SM. Nutritional regulation of muscle protein
 530 synthesis with resistance exercise: strategies to enhance anabolism. Nutr Metab (Lond)
 531 2012; 9: 40. Doi:10.1186/1743-7075-9-40
- [8] Teyhen DS. Rehabilitative Ultrasound Imaging: The Roadmap Ahead. J Orthop Sports Phys
 Ther 2007; 37: 431–433. Doi:10.2519/jospt.2007.0107

[9] Prentice CLS, Milanese S, Massy-Westropp N, et al. The reliability of rehabilitative ultrasound to measure lateral abdominal muscle thickness: A systematic review and metaanalysis. Musculoskeletal Science and Practice 2021; 53: 102357.

537 Doi:10.1016/j.msksp.2021.102357

- 540 [11] Devorski L, Skibski A, Mangum LC. Muscle Function Obtained with Motion Mode Ultrasound and Surface Electromyography during Core Endurance Exercise. . Im Internet: https://app.jove.com/t/64335/muscle-function-obtained-with-motion-mode-ultrasoundsurface; Stand: 14.08.2023 545 [12] Skibski A, Devorski L, Mangum LC. Providing Visual Biofeedback Using Brightness 546 Mode Ultrasound During a Golf Swing. Jove 2022; e64333. Doi:10.3791/64333 [13] Mangum LC, Sutherlin MA, Saliba SA, et al. Reliability of Ultrasound Imaging Measures 547 of Transverse Abdominis and Lumbar Multifidus in Various Positions. PM&R 2016; 8: 548 549 340-347. Doi:10.1016/j.pmrj.2015.09.015 550 [14] dejong AF, Mangum LC, Hertel J. Ultrasound Imaging of the Gluteal Muscles During the 551 Y-Balance Test in Individuals With or Without Chronic Ankle Instability. J Athl Train 552 2020; 55: 49-57. Doi:10.4085/1062-6050-363-18 [15] Teyhen DS, Miltenberger CE, Deiters HM, et al. The Use of Ultrasound Imaging of the 553 554 Abdominal Drawing-in Maneuver in Subjects With Low Back Pain. J Orthop Sports Phys 555 Ther 2005; 35: 346–355. Doi:10.2519/jospt.2005.35.6.346 [16] Loturco I, Gil S, Laurino CF de S, et al. Differences in muscle mechanical properties 556 557 between elite power and endurance athletes: a comparative study. J Strength Cond Res 558 2015; 29: 1723-1728. Doi:10.1519/JSC.0000000000000803 559 [17] Parmar A, Scott M, Brand C, et al. An assessment of the contractile properties of the 560 shoulder musculature in elite volleyball players using tensiomyography. Int J Sports Phys 561 Ther 2020; 15: 1099–1109. Doi:10.26603/ijspt20201099 562 [18] García-García O, Cancela-Carral JM, Huelin-Trillo F. Neuromuscular profile of top-level 563 women kayakers assessed through tensiomyography. J Strength Cond Res 2015; 29: 844– 564 853. Doi:10.1519/JSC.000000000000002 565 [19] Piqueras-Sanchiz F, Martín-Rodríguez S, Pareja-Blanco F, et al. Mechanomyographic 566 Measures of Muscle Contractile Properties are Influenced by Electrode Size and
- 567 Stimulation Pulse Duration. Sci Rep 2020; 10: 8192. Doi:10.1038/s41598-020-65111-z
- 568 [20] de Paula Simola RÁ, Harms N, Raeder C, et al. Tensiomyography reliability and prediction 569 of changes in muscle force following heavy eccentric strength exercise using muscle 570 mechanical properties. Sports Technology 2015; 8: 58–66. 571 Doi:10.1080/19346182.2015.1117475
- 572 [21] Macgregor LJ, Hunter AM, Orizio C, et al. Assessment of Skeletal Muscle Contractile 573 Properties by Radial Displacement: The Case for Tensiomyography. Sports Med 2018; 48: 574 1607-1620. Doi:10.1007/s40279-018-0912-6

- [10] Lanza MB, Rock K, Marchese V, et al. Ultrasound measures of muscle thickness and 538
- 539 subcutaneous tissue from the hip abductors: Inter- and intra-rater reliability.
 - Musculoskeletal Science and Practice 2022; 62: 102612. Doi:10.1016/j.msksp.2022.102612
- 541 542 543 544

575 576 577	[22]	Collett J, Dawes H, Meaney A, et al. Exercise for multiple sclerosis: a single-blind randomized trial comparing three exercise intensities. Mult Scler 2011; 17: 594–603. Doi:10.1177/1352458510391836
578 579 580	[23]	Wens I, Dalgas U, Vandenabeele F, et al. High Intensity Exercise in Multiple Sclerosis: Effects on Muscle Contractile Characteristics and Exercise Capacity, a Randomised Controlled Trial. Plos One 2015; 10: e0133697. Doi:10.1371/journal.pone.0133697
581 582 583	[24]	Keytsman C, Hansen D, Wens I, et al. Impact of high-intensity concurrent training on cardiovascular risk factors in persons with multiple sclerosis - pilot study. Disabil Rehabil 2019; 41: 430–435. Doi:10.1080/09638288.2017.1395086
584 585 586	[25]	Laurentino GC, Ugrinowitsch C, Roschel H, et al. Strength training with blood flow restriction diminishes myostatin gene expression. Med Sci Sports Exerc 2012; 44: 406–412. Doi:10.1249/MSS.0b013e318233b4bc
587 588 589 590	[26]	Vechin FC, Libardi CA, Conceição MS, et al. Comparisons between low-intensity resistance training with blood flow restriction and high-intensity resistance training on quadriceps muscle mass and strength in elderly. J Strength Cond Res 2015; 29: 1071–1076. Doi:10.1519/JSC.000000000000703
591 592 593	[27]	Schoenfeld BJ, Peterson MD, Ogborn D, et al. Effects of Low- vs. High-Load Resistance Training on Muscle Strength and Hypertrophy in Well-Trained Men. J Strength Cond Res 2015; 29: 2954–2963. Doi:10.1519/JSC.000000000000958
594 595 596	[28]	Hill EC, Housh TJ, Smith CM, et al. Acute changes in muscle thickness, edema, and blood flow are not different between low-load blood flow restriction and non-blood flow restriction. Clin Physiol Funct Imaging 2021; 41: 452–460. Doi:10.1111/cpf.12720
597 598 599	[29]	Hill EC, Housh TJ, Keller JL, et al. Early phase adaptations in muscle strength and hypertrophy as a result of low-intensity blood flow restriction resistance training. Eur J Appl Physiol 2018; 118: 1831–1843. Doi:10.1007/s00421-018-3918-8
600 601 602	[30]	Hill EC. The Effects of 4 Weeks of Blood Flow Restriction and Low-load Resistance Training on Muscle Strength, Power, Hypertrophy, and Neuromuscular Adaptation. University of Nebraska - Lincoln; 2019
603 604 605	[31]	Hill EC. Eccentric, but not concentric blood flow restriction resistance training increases muscle strength in the untrained limb. Phys Ther Sport 2020; 43: 1–7. Doi:10.1016/j.ptsp.2020.01.013
606 607 608	[32]	Mattar MA, Gualano B, Perandini LA, et al. Safety and possible effects of low-intensity resistance training associated with partial blood flow restriction in polymyositis and dermatomyositis. Arthritis Res Ther 2014; 16: 473. Doi:10.1186/s13075-014-0473-5

[33] Lamberti N, Straudi S, Donadi M, et al. Effectiveness of blood flow-restricted slow walking
on mobility in severe multiple sclerosis: A pilot randomized trial. Scand J Med Sci Sports
2020; 30: 1999–2009. Doi:10.1111/sms.13764

- [34] Ogawa H, Nakajima T, Shibasaki I, et al. Low-Intensity Resistance Training with Moderate 612 613 Blood Flow Restriction Appears Safe and Increases Skeletal Muscle Strength and Size in 614 Cardiovascular Surgery Patients: A Pilot Study. J Clin Med 2021; 10: 547. 615 Doi:10.3390/jcm10030547 616 [35] Takarada Y, Takazawa H, Ishii N. Applications of vascular occlusion diminish disuse atrophy of knee extensor muscles. Med Sci Sports Exerc 2000; 32: 2035–2039. 617 618 Doi:10.1097/00005768-200012000-00011 619 [36] Hughes L, Rosenblatt B, Haddad F, et al. Comparing the Effectiveness of Blood Flow 620 Restriction and Traditional Heavy Load Resistance Training in the Post-Surgery 621 Rehabilitation of Anterior Cruciate Ligament Reconstruction Patients: A UK National 622 Health Service Randomised Controlled Trial. Sports Med 2019; 49: 1787–1805. 623 Doi:10.1007/s40279-019-01137-2 [37] Abe T, Sakamaki M, Fujita S, et al. Effects of low-intensity walk training with restricted leg 624 625 blood flow on muscle strength and aerobic capacity in older adults. J Geriatr Phys Ther 626 2010; 33: 34-40 [38] Yasuda T, Fukumura K, Iida H, et al. Effect of low-load resistance exercise with and 627 628 without blood flow restriction to volitional fatigue on muscle swelling. Eur J Appl Physiol 629 2015; 115: 919-926. Doi:10.1007/s00421-014-3073-9 630 [39] Barbalho M, Rocha AC, Seus TL, et al. Addition of blood flow restriction to passive 631 mobilization reduces the rate of muscle wasting in elderly patients in the intensive care unit: 632 a within-patient randomized trial. Clin Rehabil [40] Proppe CE, Aldeghi TM, Rivera PM, et al. 75-repetition versus sets to failure of blood flow 633 634 restriction exercise on indices of muscle damage in women. Eur J Sport Sci 2023; 1–9. 635 Doi:10.1080/17461391.2023.2201813 636 [41] Gray SM, Cuomo AM, Proppe CE, et al. Effects of Sex and Cuff Pressure on Physiological 637 Responses during Blood Flow Restriction Resistance Exercise in Young Adults. Med Sci 638 Sports Exerc 2023; 55: 920–931. Doi:10.1249/MSS.000000000003103
- [42] Loenneke JP, Allen KM, Mouser JG, et al. Blood flow restriction in the upper and lower
 limbs is predicted by limb circumference and systolic blood pressure. Eur J Appl Physiol
 2015; 115: 397–405. Doi:10.1007/s00421-014-3030-7
- [43] Reis JF, Fatela P, Mendonca GV, et al. Tissue Oxygenation in Response to Different
 Relative Levels of Blood-Flow Restricted Exercise. Front Physiol 2019; 10: 407.
 Doi:10.3389/fphys.2019.00407
- [44] Weir JP. Quantifying test-retest reliability using the intraclass correlation coefficient and
 the SEM. J Strength Cond Res 2005; 19: 231–240. Doi:10.1519/15184.1

- 647 [45] Hughes L, Jeffries O, Waldron M, et al. Influence and reliability of lower-limb arterial648 occlusion pressure at different body positions. Peerj 2018; 6: e4697.
- 649 Doi:10.7717/peerj.4697
- [46] Hunt JEA, Stodart C, Ferguson RA. The influence of participant characteristics on the
 relationship between cuff pressure and level of blood flow restriction. Eur J Appl Physiol
 2016; 116: 1421–1432. Doi:10.1007/s00421-016-3399-6
- [47] Aniceto RR, da Silva Leandro L. Practical Blood Flow Restriction Training: New
 Methodological Directions for Practice and Research. Sports Med Open 2022; 8: 87.
 Doi:10.1186/s40798-022-00475-2
- [48] FDA-NIH Biomarker Working Group. BEST (Biomarkers, endpoints, and other Tools)
 Resource. Silver Spring (MD): Food and Drug Administration (US); 2016
- [49] Califf RM. Biomarker definitions and their applications. Exp Biol Med (Maywood) 2018;
 243: 213–221. Doi:10.1177/1535370217750088
- [50] Gong F, Wei H-X, Li Q, et al. Evaluation and Comparison of Serological Methods for
 COVID-19 Diagnosis. Front Mol Biosci 2021; 8: 682405. Doi:10.3389/fmolb.2021.682405
- 662 [51] Katsanos AH, Kyriakidi K, Karassa FB, et al. Biomarker Development in Chronic663 Inflammatory Diseases. 2017
- [52] Haro I, Sanmartí R, Gómara MJ. Implications of Post-Translational Modifications in
 Autoimmunity with Emphasis on Citrullination, Homocitrullination and Acetylation for the
 Pathogenesis, Diagnosis and Prognosis of Rheumatoid Arthritis. Int J Mol Sci 2022; 23:
 15803. Doi:10.3390/ijms232415803
- [53] Fosang AJ, Last K, Stanton H, et al. Neoepitope antibodies against MMP-cleaved and
 aggrecanase-cleaved aggrecan. Methods Mol Biol 2010; 622: 312–347. Doi:10.1007/978-160327-299-5_19
- 671 [54] Rogers LD, Overall CM. Proteolytic post-translational modification of proteins: proteomic
 672 tools and methodology. Mol Cell Proteomics 2013; 12: 3532–3542.
 673 Doi:10.1074/mcp.M113.031310
- 674 [55] Arvanitidis A, Henriksen K, Karsdal MA, et al. Neo-epitope Peptides as Biomarkers of
 675 Disease Progression for Muscular Dystrophies and Other Myopathies. J Neuromuscul Dis
 676 2016; 3: 333–346. Doi:10.3233/JND-160150
- [56] Karsdal MA, Henriksen K, Leeming DJ, et al. Novel combinations of Post-Translational
 Modification (PTM) neo-epitopes provide tissue-specific biochemical markers--are they the
- cause or the consequence of the disease? Clin Biochem 2010; 43: 793–804.
- 680 Doi:10.1016/j.clinbiochem.2010.03.015

- [57] Drescher C, Konishi M, Ebner N, et al. Loss of muscle mass: Current developments in cachexia and sarcopenia focused on biomarkers and treatment. Int J Cardiol 2016; 202:
 766–772. Doi:10.1016/j.ijcard.2015.10.033
- [58] Nedergaard A, Karsdal MA, Sun S, et al. Serological muscle loss biomarkers: an overview
 of current concepts and future possibilities. J Cachexia Sarcopenia Muscle 2013; 4: 1–17.
 Doi:10.1007/s13539-012-0086-2
- [59] Nedergaard A, Sun S, Karsdal MA, et al. Type VI collagen turnover-related peptides-novel
 serological biomarkers of muscle mass and anabolic response to loading in young men. J
 Cachexia Sarcopenia Muscle 2013; 4: 267–275. Doi:10.1007/s13539-013-0114-x
- [60] Nedergaard A, Dalgas U, Primdahl H, et al. Collagen fragment biomarkers as serological
 biomarkers of lean body mass a biomarker pilot study from the DAHANCA25B cohort
 and matched controls. J Cachexia Sarcopenia Muscle 2015; 6: 335–342.
 Doi:10.1002/jcsm.12027

694 [61] Sun S, Henriksen K, Karsdal MA, et al. Measurement of a MMP-2 degraded Titin fragment
695 in serum reflects changes in muscle turnover induced by atrophy. Exp Gerontol 2014; 58:
696 83–89. Doi:10.1016/j.exger.2014.07.016

- [62] Nielsen MJ, Nedergaard AF, Sun S, et al. The neo-epitope specific PRO-C3 ELISA
 measures true formation of type III collagen associated with liver and muscle parameters.
 Am J Transl Res 2013; 5: 303–315
- [63] Reule CA, Scholz C, Schoen C, et al. Reduced muscular fatigue after a 12-week leucinerich amino acid supplementation combined with moderate training in elderly: a randomised,
 placebo-controlled, double-blind trial. BMJ Open Sport Exerc Med 2016; 2: e000156.
 Doi:10.1136/bmjsem-2016-000156
- [64] Monti E, Sarto F, Sartori R, et al. C-terminal agrin fragment as a biomarker of muscle
 wasting and weakness: a narrative review. J Cachexia Sarcopenia Muscle 2023; 14: 730–
 744. Doi:10.1002/jcsm.13189
- Thambisetty M, Lovestone S. Blood-based biomarkers of Alzheimer's disease: challenging
 but feasible. Biomark Med 2010; 4: 65–79. Doi:10.2217/bmm.09.84
- [66] Goodpaster BH, Park SW, Harris TB, et al. The loss of skeletal muscle strength, mass, and
 quality in older adults: the health, aging and body composition study. J Gerontol A Biol Sci
 Med Sci 2006; 61: 1059–1064. Doi:10.1093/gerona/61.10.1059
- [67] Russ DW, Gregg-Cornell K, Conaway MJ, et al. Evolving concepts on the age-related
 changes in "muscle quality". J Cachexia Sarcopenia Muscle 2012; 3: 95–109.
 Doi:10.1007/s13539-011-0054-2
- [68] Delmonico MJ, Harris TB, Visser M, et al. Longitudinal study of muscle strength, quality,
 and adipose tissue infiltration. Am J Clin Nutr 2009; 90: 1579–1585.
 Doi:10.3945/ajcn.2009.28047

- [69] Di Girolamo FG, Fiotti N, Milanović Z, et al. The Aging Muscle in Experimental Bed Rest:
 A Systematic Review and Meta-Analysis. Front Nutr 2021; 8: 633987.
- 720 Doi:10.3389/fnut.2021.633987
- [70] maclennan RJ, Sahebi M, Becker N, et al. Declines in skeletal muscle quality vs. Size
 following two weeks of knee joint immobilization. Peerj 2020; 8: e8224.
 Doi:10.7717/peerj.8224
- [71] maclennan RJ, Ogilvie D, mcdorman J, et al. The time course of neuromuscular impairment
 during short-term disuse in young women. Physiol Rep 2021; 9: e14677.
 Doi:10.14814/phy2.14677
- [72] Correa-de-Araujo R, Harris-Love MO, Miljkovic I, et al. The Need for Standardized
 Assessment of Muscle Quality in Skeletal Muscle Function Deficit and Other AgingRelated Muscle Dysfunctions: A Symposium Report. Front Physiol 2017; 8: 87.
 Doi:10.3389/fphys.2017.00087
- [73] Stock MS, Thompson BJ. Echo intensity as an indicator of skeletal muscle quality:
 applications, methodology, and future directions. Eur J Appl Physiol 2021; 121: 369–380.
 Doi:10.1007/s00421-020-04556-6
- [74] Pinto MD, Silveira Pinto R, Nosaka K, et al. Do Intramuscular Temperature and Fascicle
 Angle Affect Ultrasound Echo Intensity Values? Med Sci Sports Exerc 2023; 55: 740–750.
 Doi:10.1249/MSS.0000000003082
- 737 [75] Arts IMP, Schelhaas HJ, Verrijp KCN, et al. Intramuscular fibrous tissue determines
 738 muscle echo intensity in amyotrophic lateral sclerosis. Muscle Nerve 2012; 45: 449–450.
 739 Doi:10.1002/mus.22254
- [76] Pillen S, Tak RO, Zwarts MJ, et al. Skeletal Muscle Ultrasound: Correlation Between
 Fibrous Tissue and Echo Intensity. Ultrasound in Medicine & Biology 2009; 35: 443–446.
 Doi:10.1016/j.ultrasmedbio.2008.09.016
- [77] Young H-J, Jenkins NT, Zhao Q, et al. Measurement of intramuscular fat by muscle echo
 intensity. Muscle Nerve 2015; 52: 963–971. Doi:10.1002/mus.24656
- [78] Tanaka NI, Ogawa M, Yoshiko A, et al. Validity of Extended-Field-of-View Ultrasound
 Imaging to Evaluate Quantity and Quality of Trunk Skeletal Muscles. Ultrasound Med Biol
 2021; 47: 376–385. Doi:10.1016/j.ultrasmedbio.2020.11.006
- [79] Stock MS, Mota JA, Hernandez JM, et al. Echo intensity and muscle thickness as predictors
 Of athleticism and isometric strength in middle-school boys. Muscle Nerve 2017; 55: 685–
 692. Doi:10.1002/mus.25395
- [80] Komforti D, Joffe C, Magras A, et al. Does skeletal muscle morphology or functional
 performance better explain variance in fast gait speed in older adults? Aging Clin Exp Res
 2021; 33: 921–931. Doi:10.1007/s40520-020-01596-y

- [81] Stock MS, Oranchuk DJ, Burton AM, et al. Age-, sex-, and region-specific differences in
 skeletal muscle size and quality. Appl Physiol Nutr Metab 2020; 45: 1253–1260.
 Doi:10.1139/apnm-2020-0114
- [82] Wong V, Spitz RW, Bell ZW, et al. Exercise induced changes in echo intensity within the muscle: a brief review. J Ultrasound 2020; 23: 457–472. Doi:10.1007/s40477-019-00424-y
- [83] Dankel SJ, Abe T, Bell ZW, et al. The Impact of Ultrasound Probe Tilt on Muscle
 Thickness and Echo-Intensity: A Cross-Sectional Study. J Clin Densitom 2020; 23: 630–
 638. Doi:10.1016/j.jocd.2018.10.003
- [84] Varanoske A, Coker N, Johnson B-A, et al. Effects of Rest Position on Morphology of the
 Vastus Lateralis and Its Relationship with Lower-Body Strength and Power. JFMK 2019; 4:
 64. Doi:10.3390/jfmk4030064
- [85] Carr JC, Gerstner GR, Voskuil CC, et al. The Influence of Sonographer Experience on
 Skeletal Muscle Image Acquisition and Analysis. J Funct Morphol Kinesiol 2021; 6: 91.
 Doi:10.3390/jfmk6040091
- [86] Girts RM, Harmon KK, Pagan JI, et al. The influence of ultrasound image depth and gain
 on skeletal muscle echo intensity. Appl Physiol Nutr Metab 2022; 47: 839–846.
 Doi:10.1139/apnm-2021-0810
- [87] Seynnes OR, Cronin NJ. Simple Muscle Architecture Analysis (SMA): An imagej macro
 tool to automate measurements in B-mode ultrasound scans. PLOS ONE 2020; 15:
 e0229034. Doi:10.1371/journal.pone.0229034
- [88] Ritsche P, Wirth P, Franchi MV, et al. Acsauto-semi-automatic assessment of human vastus
 lateralis and rectus femoris cross-sectional area in ultrasound images | Scientific Reports.
 Sci Rep
- [89] Medicine AC of S, Lohman T, Milliken LA. ACSM's Body Composition Assessment.
 Human Kinetics; 2019
- [90] Kasper AM, Langan-Evans C, Hudson JF, et al. Come Back Skinfolds, All Is Forgiven: A
 Narrative Review of the Efficacy of Common Body Composition Methods in Applied
 Sports Practice. Nutrients 2021; 13: 1075. Doi:10.3390/nu13041075
- [91] Santos DA, Dawson JA, Matias CN, et al. Reference Values for Body Composition and
 Anthropometric Measurements in Athletes. Plos ONE 2014; 9: e97846.
 Doi:10.1371/journal.pone.0097846
- 785 [92] Fukuda D. Assessments for Sport and Athletic Performance. Human Kinetics; 2019
- [93] Slater G, Woolford S, Sport AI of, et al. Assessment of Physique. In: Physiological Tests
 for Elite Athletes. Human Kinetics; 2013

- 788 [94] Bonilla DA, De León LG, Alexander-Cortez P, et al. Simple anthropometry-based 789 calculations to monitor body composition in athletes: Scoping review and reference values. 790 Nutr Health 2022; 28: 95-109. Doi:10.1177/02601060211002941 791 [95] Defreitas JM, Beck TW, Stock MS, et al. A comparison of techniques for estimating 792 training-induced changes in muscle cross-sectional area. J Strength Cond Res. 793 Doi:10.1519/JSC.0b013e3181ec86f3 [96] Moritani T, devries HA. Neural factors versus hypertrophy in the time course of muscle 794 795 strength gain - pubmed. Am J Phys Med 1979; 58: 115–130 796 [97] Housh DJ, Housh TJ, Weir JP, et al. Anthropometric estimation of thigh muscle cross-797 sectional area. Med Sci Sports Exerc 1995; 27: 784–791 798 [98] Duarte CK, De Abreu Silva L, Castro CF, et al. Prediction equations to estimate muscle mass using anthropometric data: a systematic review. Nutr Rev 2023; 81: 1414-1440. 799 800 Doi:10.1093/nutrit/nuad022 801 [99] Lukaski H, Raymond-Pope CJ. New Frontiers of Body Composition in Sport. Int J Sports Med 2021; 42: 588-601. Doi:10.1055/a-1373-5881 802 803 [100] Kim C-H, Park J-H, Kim H, et al. Modeling the human body shape in bioimpedance vector measurements. Proc IEEE Inst Electr Electron Eng 2010; 804 805 doi:10.1109/IEMBS.2010.5627664 806 [101] Di Vincenzo O, Marra M, Scalfi L. Bioelectrical impedance phase angle in sport: a 807 systematic review. Journal of the International Society of Sports Nutrition 2019; 16: 49. 808 Doi:10.1186/s12970-019-0319-2 809 [102] Cebrián-Ponce Á, Irurtia A, Carrasco-Marginet M, et al. Electrical Impedance 810 Myography in Health and Physical Exercise: A Systematic Review and Future Perspectives. 811 Front Physiol 2021; 12 [103] Terminology | International Association for the Study of Pain. . Im Internet: 812 813 https://www.iasp-pain.org/resources/terminology/; Stand: 13.11.2023 [104] Cao Q-W, Peng B-G, Wang L, et al. Expert consensus on the diagnosis and treatment of 814 815 myofascial pain syndrome. World J Clin Cases 2021; 9: 2077–2089. 816 Doi:10.12998/wjcc.v9.i9.2077 817 [105] Harden RN, Bruehl SP, Gass S, et al. Signs and symptoms of the myofascial pain 818 syndrome: a national survey of pain management providers. Clin J Pain 2000; 16: 64–72.
- 819 Doi:10.1097/00002508-200003000-00010
- [106] Li L, Stoop R, Clijsen R, et al. Criteria Used for the Diagnosis of Myofascial Trigger
 Points in Clinical Trials on Physical Therapy: Updated Systematic Review. Clin J Pain
 2020; 36: 955–967. Doi:10.1097/AJP.00000000000875

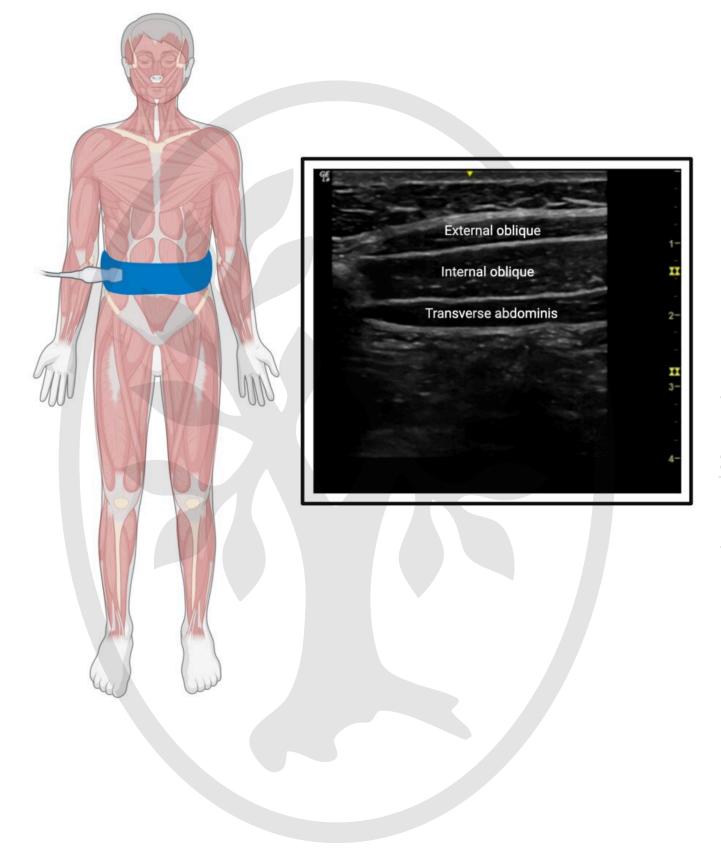
- 823 [107] Fernández-de-Las-Peñas C, Dommerholt J. International Consensus on Diagnostic
 824 Criteria and Clinical Considerations of Myofascial Trigger Points: A Delphi Study. Pain
 825 Med 2018; 19: 142–150. Doi:10.1093/pm/pnx207
- 826 [108] Simons DG, Stolov WC. Microscopic features and transient contraction of palpable
 827 bands in canine muscle. Am J Phys Med 1976; 55: 65–88
- [109] Srbely JZ, Dickey JP, Bent LR, et al. Capsaicin-Induced Central Sensitization Evokes
 Segmental Increases in Trigger Point Sensitivity in Humans. J Pain 2010; 11: 636–643.
 Doi:10.1016/j.jpain.2009.10.005
- [110] Lucas N, Macaskill P, Irwig L, et al. Reliability of physical examination for diagnosis of
 myofascial trigger points: a systematic review of the literature. Clin J Pain 2009; 25: 80–89.
 Doi:10.1097/AJP.0b013e31817e13b6
- [111] Wytrążek M, Huber J, Lipiec J, et al. Evaluation of Palpation, Pressure Algometry, and
 Electromyography for Monitoring Trigger Points in Young Participants. J Manipulative
 Physiol Ther 2015; 38: 232–243. Doi:10.1016/j.jmpt.2014.12.005
- [112] Oliveira AK de, Dibai-Filho AV, Soleira G, et al. Reliability of pressure pain threshold
 on myofascial trigger points in the trapezius muscle of women with chronic neck pain. Rev
 Assoc Med Bras (1992) 2021; 67: 708–712. Doi:10.1590/1806-9282.20201149
- 840 [113] Mazza DF, Boutin RD, Chaudhari AJ. Assessment of Myofascial Trigger Points via
 841 Imaging: A Systematic Review. Am J Phys Med Rehabil 2021; 100: 1003–1014.
 842 Doi:10.1097/PHM.0000000001789
- 843 [114] Sikdar S, Shah JP, Gebreab T, et al. Novel applications of ultrasound technology to
 844 visualize and characterize myofascial trigger points and surrounding soft tissue. Arch Phys
 845 Med Rehabil 2009; 90: 1829–1838. Doi:10.1016/j.apmr.2009.04.015
- [115] Luiza da Silva Queiroz M, Bezerra Diniz PR, Nepomuceno Montenegro EJ, et al. MRI in
 migraineurs: are there abnormalities in the area where the myofascial trigger points are
 palpable and in volume measurements? J Bodyw Mov Ther 2020; 24: 260–266.
 Doi:10.1016/j.jbmt.2020.02.025

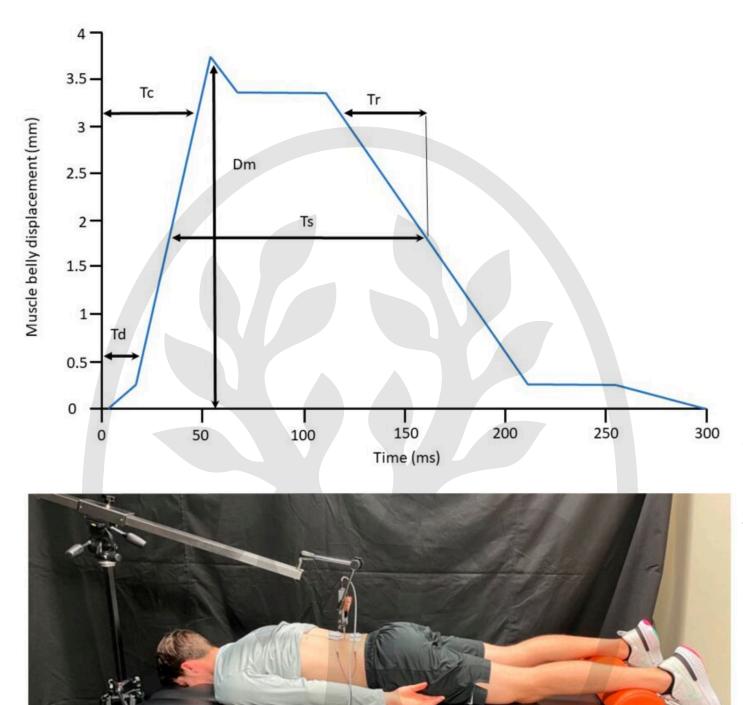
850 Figure Legends

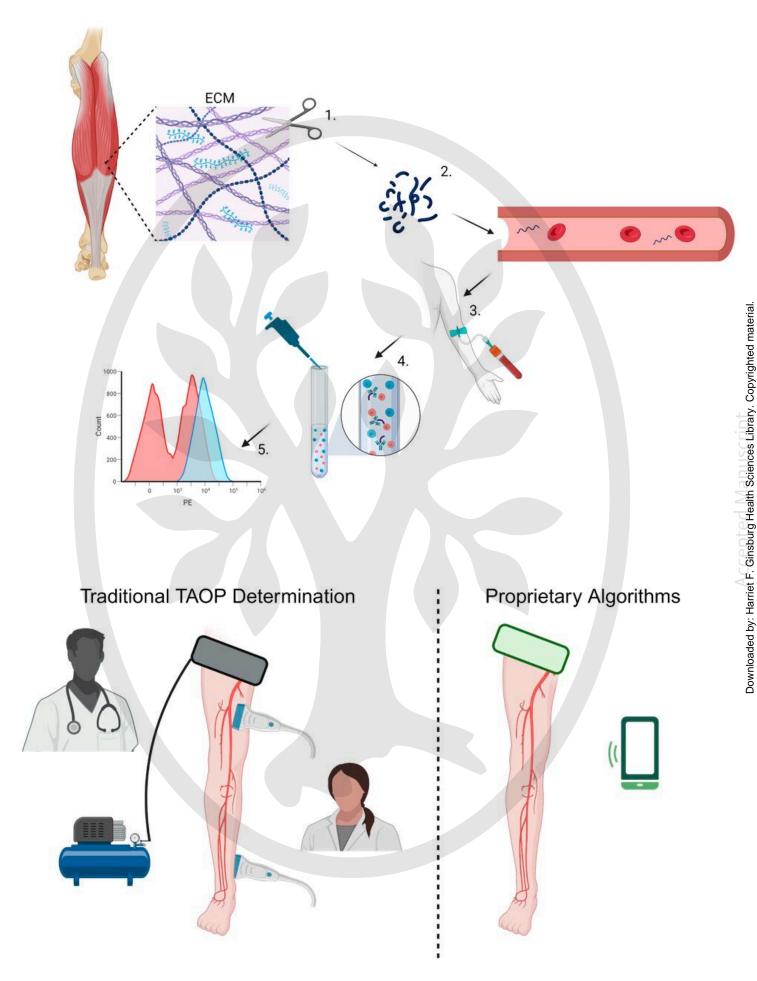
- Figure 1. Dynamic imaging (B-mode) of lateral abdominal wall musculature using an elastic belt to keeplinear transducer fixed to the abdomen throughout movement. Created with BioRender.com.
- Figure 2. Muscle belly displacement versus time the phases of contraction elicited by TMG with key
- 855 parameters comprise displacement (Dm), contraction time (Tc), delay time (Td), contraction velocity (Vc)
- 856 (Vc=[90%Dm-10%Dm/Tc]), sustain time (Ts), and half-relaxation time (Tr). (Top). An example of
- 857 (TMG) placement for the lumbar erector spinae (bottom).
- 858 Figure 3. Illustrates the gold standard for the determination of TAOP which requires at least one
- technician. More recently, innovative approaches have been developed which leverage proprietary
- algorithms to determine TAOP and do not require a trained technician(s). These devices can often be

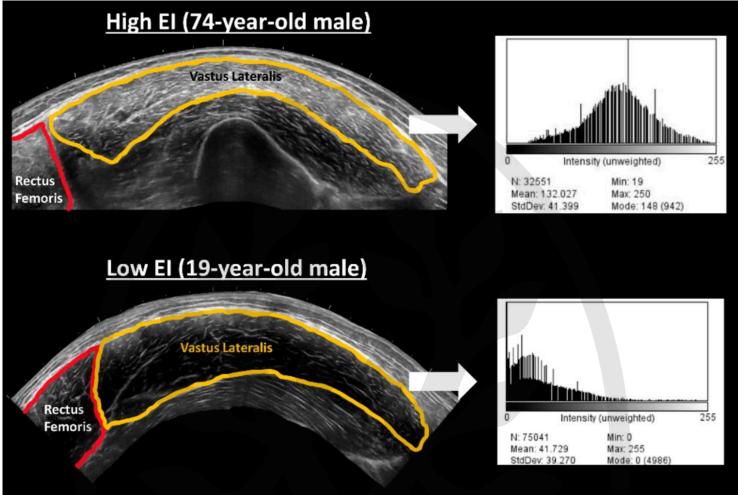
- 861 controlled wirelessly from a mobile app and detached from the air source facilitating, greater utility in862 exercise and rehabilitation settings. Created with BioRender.com.
- 863 Figure 4. Detection of serological peptide fragments. 1. Proteolytic cleavage of collagen protein 2.
- Peptide fragments enter blood stream, 3. Blood sample obtained, 4. Antibodies raised against neoepitope
 markers, 5. Quantification vis assay e.g., flow cytometry. Created with BioRender.com.
- Figure 5. Example B-mode ultrasound images and echo intensity (EI) analyses of the vastus lateralis for
 an older (top) and younger (bottom) male. Note the vastly different pixel distributions for the two images.
 These images are a fairly accurate depiction of published findings, as many studies have reported higher
 echo intensity among older adults.
- Figure 6. Changes in site-specific skinfold thickness and circumference (or body mass) as potential
 proxies of body fat and muscle mass, respectively. The examples provided indicate trends toward and
 away from A) muscle growth/hypertrophy, B) adiposity, C) muscle growth/hypertrophy with leanness,
 and D) muscle growth/hypertrophy with adiposity. Created with BioRender.com.
- Figure 7. Overview of Methods to Identify a Myofascial Trigger Point. Myofascial trigger points may beidentified with palpation or novel imaging techniques. Although not comprehensive, novel imaging
- techniques may include ultrasound or magnetic resonance imaging (MRI). Created with BioRender.com.

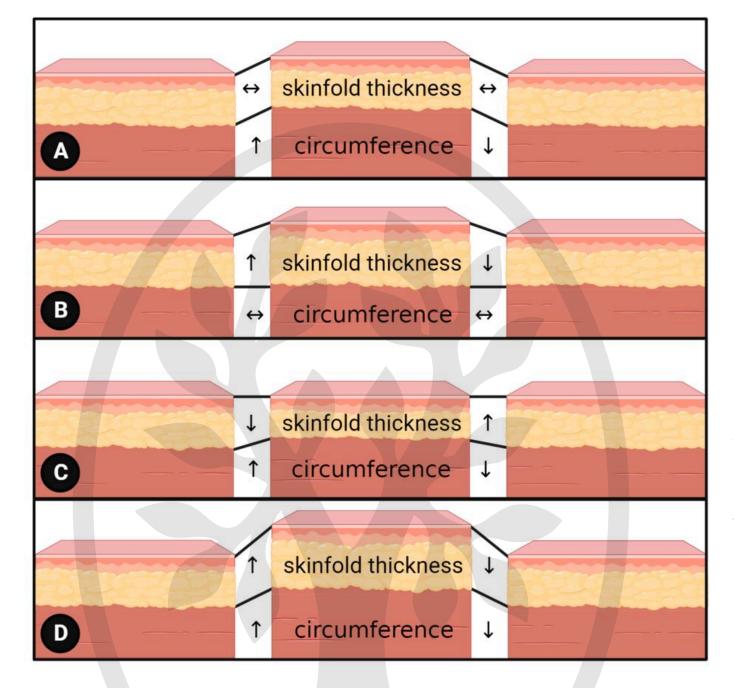


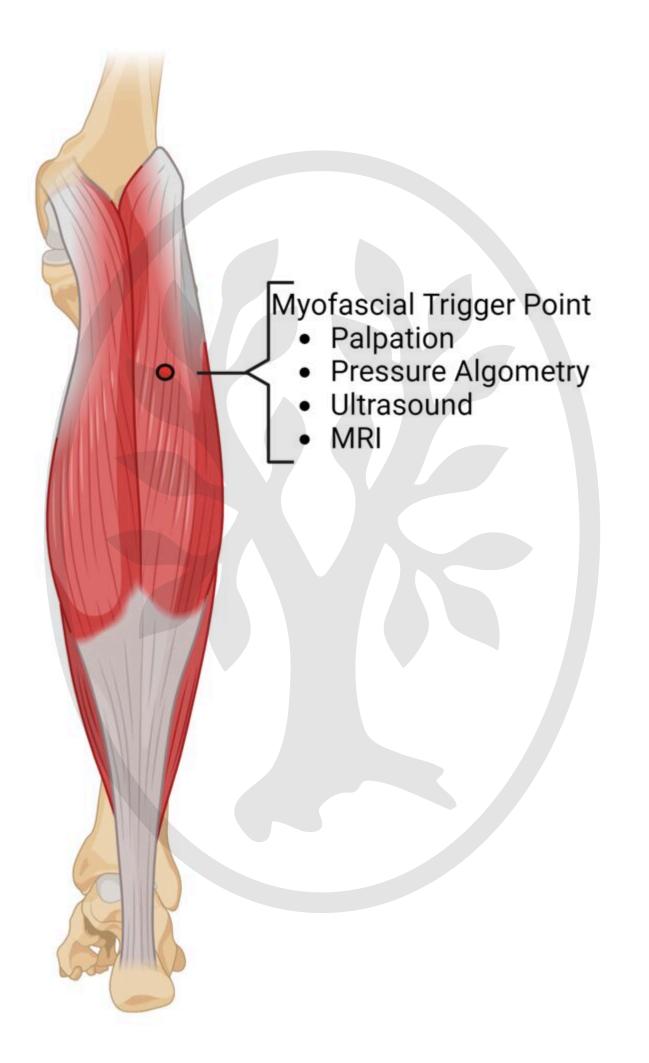












Downloaded by: Harriet F. Ginsburg Health Sciences Library. Copyrighted material.

his article is protected by copyright. All rights reserved.

